

Size Dependent Free Solution DNA Electrophoresis in Structured Micro Fluidic Systems

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Abstract

A new concept based on microstructuring techniques is presented to achieve size dependent electrophoretic migration of DNA in free solution. Topographically structured microfluidic channels with periodical cavities in the range of the radius of gyration of the tested DNA molecules ($<3\ \mu\text{m}$) are produced by moulding of polydimethylsiloxane on a master wafer fabricated by contact lithography of SU-8 photoresist. The electrophoretic migration of single DNA molecules stained with the intercalator YOYO was investigated by real-time fluorescence video microscopy. It could be demonstrated for large DNA molecules that λ -DNA (48 kbp) always migrated significantly faster than T2-DNA (164 kbp) in the two tested microchannel layouts. Integration into a microdevice for DNA separation in free solution is now in progress.

Introduction

DNA electrophoresis represents a very important tool in molecular biology and genome analysis. In respect to the length independent mobility of DNA molecules in free solution, DNA electrophoresis is commonly carried out in gels. To enhance throughput and to shorten analysis time electrophoretic separation of DNA in capillaries or on microchips has been developed. However, the transfer of DNA separation to the microchip format has only been achieved for analytical applications in distinct DNA size ranges, as for example DNA sequencing [1–5]. This is mainly due to technical problems involved with the transfer and integration of conventional technology, e.g. the incorporation of cross linked or high viscosity gels, into capillaries and microchips [6].

Fluorescence microscopy with ultra sensitive cameras allows the investigation of single (usually with fluorescent intercalators) labelled DNA molecules. This has been used by several groups to enlighten DNA migration mechanisms in cross linked [7;8] and diluted gels [8–12] as well as artificial gels composed of microarrays [13–15]. Microfabrication techniques were also involved in the development of completely new concepts for DNA migration and separation, e.g. entropic traps [16–19] or molecular ratchets [20;21].

Our work focuses on the free solution separation of DNA in microfluidic devices using the influence of topographical structuring. The selection of microfluidic systems provides a huge pool of topographical variation, which can be achieved by conventional microstructuring techniques. Microchannels were fabricated by moulding of polydimethylsiloxane (PDMS) on a master wafer providing the inverted relief of the

desired microstructures that were previously fabricated with an epoxy based photoresist (SU-8) by contact lithography. Feature sizes in the range of the radius of gyration of the DNA molecules are produced. The mobility of λ -DNA (48 kbp) and T2-DNA (164 kbp) was investigated by real-time video recording of the migration of individual DNA molecules stained with the fluorescent intercalator YOYO.

Materials and Methods

Reagents: PDMS (Sylgard 184) was purchased from Dow Corning, USA. SU-8 50 negative photoresist, thinner γ -butyrol acetone (GBL) and developer (propyleneglycolmethylether acetate) were obtained from Microresist, GER. Si-wafers (P-Type 100, doped with boron) were from CrysTec, GER. T2- and λ -DNA (164 kbp, 48.5 kbp) were obtained from Fluka, GER and YOYO from Molecular Probes, USA. TRIS buffer, β -mercapthoethanol and NaCl were obtained from Fluka, GER and ethylenediaminetetraacetic acid (EDTA) from Sigma, GER. H₂O₂ (30%) and H₂SO₄ were from Merck, GER. Water was deionised with a Milli-Q biocel (Millipore, USA). Cover glass slides (76x26 mm²) were from Menzel Gläser, GER.

DNA staining: DNA stock solutions were diluted in TRIS buffer (10 mM TRIS, 50 mM NaCl, 1 mM EDTA, 2% (v/v) β -mercapthoethanol, pH 8.3) to a final concentration of 6 pM dsDNA. YOYO was added to obtain a final dye to basepair ratio of about 1:7.5.

Fabrication of master using photoplastic SU-8: 100 ml of SU-8 50 were diluted with 32.8 mL GBL to yield SU-8 5. After 24 h still remaining bubbles could be removed by heating to 60°C for 30 min on a hot plate. Si-wafers were cleaned with a standard piranha (H₂O₂:H₂SO₄ 1:3) etching procedure, dried on a spin coater and dehumided on a hot plate at 200°C (HTZ-303D, ATV-Technologie, GER) for 20 min. The spin-coating process on a 1001/ST147 (Convac, CH) consisted of i) spreading the photoresist over the wafer at 700 rpm for 5 sec and ii) the spin-coat process at 5200 rpm for 35 sec. The prebake process was performed by heating for 3 min at 40°C, 3 min at 65°C and 5 min at 95°C. A flood exposure unit (Modell 8011, Oriel Corporation,

USA) was used to expose the photoresist through a chrome mask (DeltaMask, NL) for 3.75 sec.

For the post exposure bake, the temperature was first ramped to 40°C, then to 60°C, and finally 95°C and held for 1, 3 and 20 min, respectively. The coated wafer was now developed for 60 sec in the developer bath. Rinses with acetone and isopropanole were performed before drying in a weak stream of nitrogen. A hard bake at 200°C for 15 min followed and cooling down was performed on the hot plate.

PDMS moulding: Prepolymer and curing agent were mixed thoroughly in a ratio of 10:1, poured on the masterwafer (see Figure 1A) and fully polymerised within 3 h at 75°C. The PDMS slab was then peeled off the masterwafer and reservoir holes were punched from the structured side. A clean cover glass and a PDMS microchannel were oxidised for 5 min in an UVO cleaner (Model 42–220, Jelight, USA) and brought into contact immediately after oxidation. The channels were filled with 0.1 M NaOH which was replaced by TRIS buffer solution after 1 h.

Channel characterisation: A 20 nm thick gold layer was evaporated (MCS 10, Bal–Tec, FL) on the PDMS cast. Scanning electron microscopy (SEM) images of the structured microchannels were taken with a LEO 1538 (LEO Electron microscopy, UK) at 5 kV acceleration voltage and a JSM–880 (Jeol, JP) at 4 kV. The height of the structured masterwafer was measured via profilometry (Dektak 3030ST, Veeco, USA).

Fluorescence Video Microscopy: The fluorescence video microscopy setup consisted of an inverted microscope (Axiovert 100, Zeiss, GER) with a filterset for fluorescence observation (XF100–3, Omega, USA), a 100x oil immersion objective (Plan Neofluar NA 1.3, Zeiss, GER) a scientific interline transfer CCD–camera (Imager 3LS, LaVision, GER) and a camera software for data analysis (DaVis 5.4.4, LaVision, GER).

A mercury arc lamp (HBO50, Zeiss, GER) was used for excitation. The microfluidic device was mounted on a x/y-stage (99S008, Ludl Electronic Products, USA). A power supply (MCN 14–2000, FUG, GER) connected to two platinum electrodes was used to apply the voltage.

Data Analysis: The determination of the velocity of DNA molecules was carried out via cross correlation analysis. With the help of command language macros (DaVis) the analysis was automated. Cross correlating two subsequent images results in an intensity image, where the positions of the highest peaks represent the most probable displacements of the molecules. Several molecules with similar displacements superpose to one peak and therefore yield an averaged velocity. The histogram containing the averaged velocities of 199 data values (corresponding to 200 image correlations) was Gauss fitted.

Results and Discussion

PDMS microstructures were fabricated by moulding on a SU-8 master wafer as shown in figure 1A and explained in detail in the experimental section. The master wafer was fabricated in a way that one or two side walls contained periodical cavities (see also figure 1D for details). Two different microchannel layouts have been used in this work: A 1.5 μm channel with periodical cavities on both lateral walls of the PDMS microchannel and a 3 μm channel with only one wall structured. Figure 1B and C show SEM images of these moulded PDMS microchannels.

The chrome mask used for the SU-8 master fabrication consisted of periodical square waveform structuring. For all moulded PDMS structures, the final periodical structures were slightly smaller than defined by the chrome mask dimensions and the periodical cavities were not a square waveform. This effect is attributed to the quality of the chrome mask, to the incomplete attachment of the mask to the SU-8 during exposure as a consequence of our experimental setup and to the diffraction limit. For the 1.5 μm channels the longitudinal periodicity ($a + c$) was determined to 2.9 μm and the lengths b and d were 1.0 and 0.8 μm , respectively. For the 3 μm structure the lengths a and c resulted in 3.0 and 2.8 μm , respectively, whereas b and d in 2.7 and 2.6 μm . The height of the master wafer structures was $2.7 \pm 0.1 \mu\text{m}$ as confirmed by profilometry measurements.

The migration of λ -DNA (48 kbp) and T2-DNA (164 kbp) was investigated in structured and planar channels. DNA molecules were stained with YOYO in a basepair to YOYO ratio of 7.5:1 in TRIS buffer, containing EDTA and mercaptoethanol. Single DNA molecules could be observed by fluorescence microscopy for at least 30 sec

without any fragmentation or significant loss in fluorescence intensity under these conditions. Only for low field strength, in which the migration velocity was a few micrometer per second, the molecules were several seconds under illumination in the typically 80 μm wide observation window. Thus the fluorescence intensity and lifetime of the stained DNA molecule was always sufficient for the video microscopy experiments.

Figure 2 demonstrates the migration of a single λ -DNA molecule in a 1.5 μm channel at a field strength of 3 V/mm. The complete transit of the molecule through the 30.15 μm display window is shown (450 pixel, 6.7 μm pixel size, 100x magnification). For the visualisation of the constrictions the channel was illuminated with a white light source with the accepted disadvantage of a higher background. However, the YOYO stained DNA molecules were still detectable with sufficient brightness and S/N-ratio.

Obviously the λ -DNA molecule is in a coiled conformation filling the periodical cavities of the microchannel. The diameter of a λ -DNA molecule in free solution can be estimated from the radius of gyration $R_g = (1/3 l_p L)^{1/2}$ according to the worm like chain model [22], where l_p and L denote persistence and contour length of the molecule, respectively. For YOYO stained λ -DNA Quake et al. [23] reported an increase in persistence length from 50 nm to 66 nm and an increase in contour length from 16 μm to 22 μm . Thus, for YOYO stained DNA the conformational diameter of the coiled λ -DNA molecules was estimated to $\sim 1.4 \mu\text{m}$. Since lateral channel constrictions and diameter of the molecule are in the same dimension an influence on the conformation and the migration of the molecules can be expected, which is confirmed by the image sequence of figure 3. During its migration, the molecule was also found to be stretched spanning one period of cavities (see Figure 3, pictures 11 to 13), but at any time its

conformation was influenced by the channel constrictions. For the longer T2–DNA molecule a statistical molecular diameter of 2.6 μm is calculated with a contour length of 72 μm [24]. The influence of the constrictions on the conformation of the T2–DNA molecule can thus be expected to be stronger.

The mobilities (defined as the averaged velocity per electric field) of λ – and T2–DNA were investigated consecutively in one microchannel. In figure 3 the mobility of the two molecules in a 1.5 μm channel at increasing electric field is shown. The mobility for λ –DNA is always found significantly higher than for T2–DNA. For comparison, the free solution mobility in a planar microchannel without constrictions and with 20 x 20 μm^2 cross section was also investigated. Here, no difference in the migration velocity for the two DNA molecules could be observed (see inset of figure 3A) and a free solution mobility of $(1,84 \pm 0.03) \cdot 10^{-4} \text{ cm}^2/\text{Vs}$ was determined.

For the 3 μm channel, the channel constrictions in all three dimensions are in the range of the diameter of the T2–DNA molecule. The migration velocity in dependence of the electric field in a 3 μm channel is shown in figure 4. As in the smaller channel configuration the λ –DNA migrates significantly faster than the longer T2–DNA.

It is interesting to note that the migration velocity of the smaller λ –DNA in the two channel geometries is always faster than the larger T2–DNA molecules. This is in contrast to the case of entropic traps where the smaller molecules migrate slower than the larger [18;19]. For entropic trapping it is inevitable that the constrictions are in the dimension of the persistence length of the DNA molecules (<100 nm). The constrictions in our channels have the dimension of the diameter of the coiled DNA molecules (1 to 3 μm) and thus do not exactly correspond to entropic traps. However, our experiments clearly demonstrated that the geometrically structured microchannels

exhibit an influence on the migration velocity dependent on molecule length. The molecules are forced into conformations by the channel constrictions which influence their migration behaviour. Other effects influencing the migration properties arise from attractive interaction of the molecules with the channel walls or the inhomogeneous electric field distribution in the microchannels as indicated by preliminary finite element simulations (data not shown). The combination of these effects causes a faster migration of shorter molecules.

Conclusion

Our work clearly demonstrates an alternative way to achieve length dependent DNA migration and separation in gel free systems. PDMS microstructures with topographical structuring affects the migration of λ - and T2-DNA molecules due to conformational effects. Individual DNA molecules could be observed and their pathways through the microstructures could be followed by real-time fluorescence video microscopy. Investigating migration velocities in structured channels showed that shorter λ -DNA migrated always significantly faster than T2-DNA. The comparison with planar channels clearly demonstrated, that without constrictions a length independent mobility is detected. The retardation of the longer T2-DNA molecules in the structured channels is attributed to entropic effects caused by conformational restrictions of the molecules, to surface interactions in the microchannels and to the distribution of the electric field in the microchannels. These phenomena will be the main issue for our future investigation on single molecule migration mechanisms. In order to investigate a larger variety of DNA sizes, an optimised microchannel geometry and a complete integrated microfluidic system with injection, separation and detection is under construction.

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Figure captions:

Figure 1: A) Process diagram of microchannel fabrication. (i) Convert shaped microchannel mould is fabricated with SU-8. (ii) PDMS is poured and cured over the mould. (iii) PDMS is released from the mould. The microchannel is thus replicated into PDMS. (iv) Reservoirs are punched through PDMS. The microchannel is closed with microscope cover slide. B) SEM image of moulded 1.5 μm PDMS microchannel and C) 3 μm channel. D) The drawing describes the variables of the periodical cavities (a, b, c and d) in the microchannels.

Figure 2:

Image sequence of a λ -DNA molecule migrating in a 1.5 μm channel. The observation window corresponds to a microchannel section of 30.15 μm . The figure represents the real sequence with 40 ms exposure and readout time between each image. The molecule requires 0.8 s to transverse the total observation window.

Figure 3: Migration velocity of (?) λ -DNA and (?) T2 DNA A) in a 1.5 μm and B) in a 3 μm channel in dependence of the applied electric field. The λ -DNA molecules migrate significantly faster than T2-DNA for field strengths higher than 2 V/mm. The dotted lines represent a least square linear fit of the data points. The inset of figure 3A shows the migration velocity of these two DNA types in a microchannel without periodical structuring (20 x 20 μm^2 cross section). The size dependence for the two molecules is lost.

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Figure 1:

T. Duong

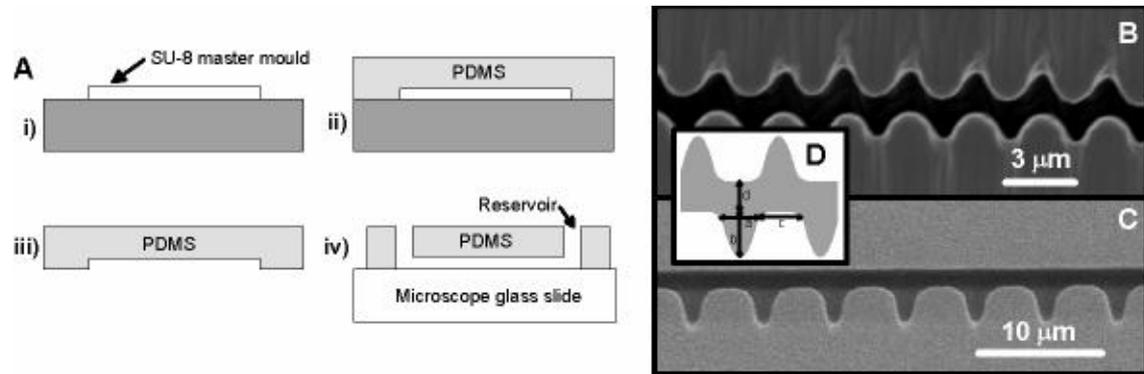


Figure 2:

T. Duong

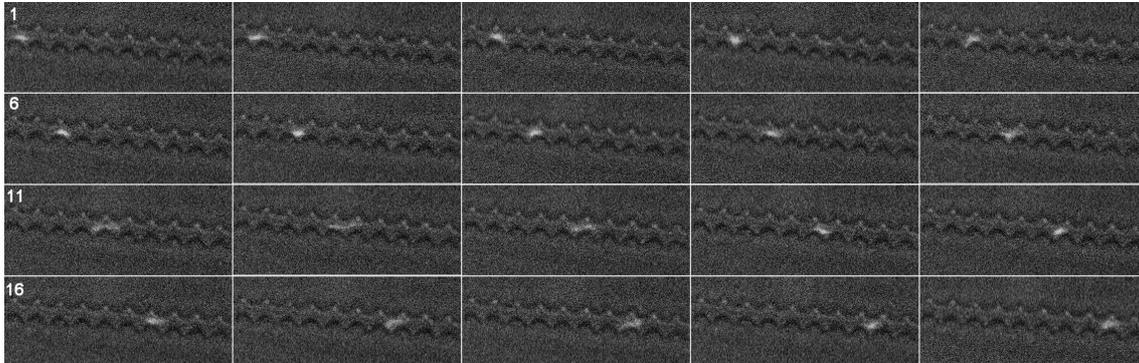


Figure 3

T. Duong

